Effects of pulmonary metabolites of prostaglandins E_2 and $F_{2\alpha}$ on guinea-pig respiratory tract

PIA YONG LO

Department of Obstetrics and Gynaecology, University of Singapore, Kandang Kerbau Hospital, Singapore 8

The effects of three pulmonary metabolites of prostaglandins E_2 and $F_{2\alpha}$ on the guinea-pig tracheal muscle *in vitro* and on lung resistance *in vivo* were investigated. Prostaglandin $F_{2\alpha}$, 13, 14-dihydro PGF_{2α} and 13,14-dihydro 15-oxo PGF_{2α} produced contractions on the tracheal muscle *in vitro*. 15-oxo PGF_{2α} relaxed some tracheal preparations but stimulated others, the stimulant action having a threshold dose range some 10-25 times lower than the relaxant doses. Prostaglandin $F_{2\alpha}$ and 13,14-dihydro PGF_{2α} increased lung resistance in anaesthetized guinea-pigs. Prostaglandin E_2 and its three pulmonary metabolites produced relaxation of guinea pig trachea *in vitro* and decreased lung resistance *in vivo*. All three metabolites were less active than PGE₂ in both the systems. The *in vitro* effects of PGF_{2α} and its metabolites were selectively blocked by polyphloretin phosphate.

Postaglandins E_2 and $F_{2\alpha}$ are present in pulmonary tissues (Änggård, 1965; Karim, Sandler & Williams, 1967) and the lungs are also involved in the metabolism of these prostaglandins. The pulmonary metabolites as well as the parent prostaglandins have pharmacological effects on airway smooth muscle. Generally, though not invariably, prostaglandins of the E series relax and those of the F series contract smooth muscle of the respiratory tract (see Smith, 1976 for references).

Dawson, Lewis & others (1974) reported that 15-oxo PGF_{2 α} is more potent than PGF_{2 α} in contracting guinea-pig tracheal and human bronchial muscles *in vitro*. However, on various airway and lung dynamics parameters in intact anaesthetized dogs, 15-oxo PGF_{2 α} was relatively inactive compared to PGF_{2 α} (Wasserman, 1975).

There is also a controversy regarding the potencies of the pulmonary metabolites of PGE₂ on bronchial smooth muscle. Boot, Dawson & Harvey (1976) found 15-oxo PGE₂, 13,14-dihydro PGE₂ and 13,14-dihydro 15-oxo PGE₂ to be less active than PGE₂ in relaxing respiratory tract smooth muscle *in vitro*. In contrast Crutchley & Piper (1975) found 15-oxo PGE₂ to be 1–1.8 times more potent than PGE₂ and the other two metabolites as active as PGE₂ in relaxing guinea-pig trachea *in vitro*. The effect of the pulmonary metabolites of PGE₂ and PGF_{2α} on lung resistance in guinea-pigs has not been investigated before.

In view of limited studies on the effects of prostaglandin metabolites on respiratory tract smooth muscles and the reported differences in the results, I have investigated the effects of three pulmonary metabolites of PGE_2 and $PGF_{2\alpha}$ on the

guinea-pig tracheal chain preparation in vitro and on guinea-pig lung resistance in vivo.

MATERIALS AND METHODS

In vitro preparation

Guinea-pig tracheal chains were prepared according to the method of Akcasu (1952). Each chain, containing 4 to 5 rings of trachea was suspended in a 25 ml organ bath containing Krebs-Ringer solution of the following composition (g litre⁻¹): sodium chloride 6·9; potassium chloride 0·35; calcium chloride ($2H_2O$) 0·37; potassium dihydrogen orthophosphate 0·14; magnesium sulphate ($7H_2O$) 0·11; glucose 1·98; sodium bicarbonate 2·1. The bathing fluid was maintained at 37° and aerated with 5% carbon dioxide in oxygen. The pH of the bath fluid was 7·4.

A load of 0.3 g was applied to the tissue and allowed to equilibrate for an hour before testing the effects of prostaglandins. Isometric contractions or relaxations were measured using a force tension transducer connected to a recording device. The dose cycle ranged from 25-40 min and contact time from 7 to 12 min. The log dose response was exercised for all compounds. Suitable submaximal doses of the metabolites of PGE₂ and PGF_{2α} were compared with similar doses of respective parent compounds using a four point bioassay method.

Effect of polyphloretin phosphate

Polyphloretin phosphate (PPP), a selective antagonist of prostaglandins (Eakins, 1972) was tested for its action on the effects of prostaglandin metabolites on guinea-pig tracheal chain preparation *in vitro*. PPP was added to the bath 1 min before the addition the agonist. Carbachol or histamine served as controls.

n vivo preparation

modification of James' (1969) method for reasuring lung resistance in anaesthetized animals was used. Guinea-pigs, 300-400 g, were anaesthewith intraperitoneal injections of 30 mg kg⁻¹ pentobarbitone sodium. A ventilation cannula was inserted into the trachea and connected to a Palmer miniature ideal pump, set at 74 strokes min⁻¹ and stroke volume of 3.5 ml. A side-arm of the outnow was connected to a non-return water valve. et at a pressure of 10 cm water. Intrapulmonary pressure was measured by a pressure transducer connected to the outflow. The jugular vein was cannulated for intravenous administration of drugs. The carotid artery was cannulated and connected to another pressure transducer for simultaneous measurement of blood pressure for monitoring the state of the animal. An interval of 10-15 min was allowed between doses. All drugs were made up to the required final dilution in 0.9% sodium chloride chloride solution. The relaxant effects of PGE₂ and metabolites were monitored indirectly by measuring the differences in increase of airway resistance to standard doses of histamine before and 1 min after intravenous injection of prostaglandins.

RESULTS

Guinea-pig tracheal chain preparations in vitro

The potencies of six pulmonary metabolites were compared with their respective parent compounds, PGE_2 or $F_{2\alpha}$. $PGF_{2\alpha}$, 13,14-dihydro $PGF_{2\alpha}$ and 13,14-dihydro 15-oxo $PGF_{2\alpha}$ produced dose-related contractions of guinea-pig trachea. 15-oxo $PGF_{2\alpha}$ in three out of six experiments relaxed the smooth muscles and in the remainder three contracted. Threshold doses for the relaxant effect ranged between 0.2 and $1.32 \,\mu g \, ml^{-1}$ whilst that for the stimulant action varied between 0.008 and 0.032 $\mu g \, ml^{-1}$. 13,14-Dihydro $PGF_{2\alpha}$ was about as potent as $PGF_{2\alpha}$ and 13,14-dihydro 15-oxo $PGF_{2\alpha}$ was the least active metabolite (Table 1).

 PGE_2 , 15-oxo PGE_2 , 13,14-dihydro PGE_2 and 13,14-dihydro 15-keto PGE_2 relaxed the guinea-pig tracheal muscles. All three metabolites were much less active than PGE_2 (Table 1). Sometimes PGE_2 and its metabolites caused an initial transient contraction (cf. Sweatman & Collier, 1968).

Effect of polyphloretin phosphate (PPP)

Over a wide range of concentrations (0.8 to $400 \mu g$

Table 1. Relative potencies of $PGF_{2\alpha}$ and PGE_2 metabolites on guinea-pig tracheal chain preparation in vitro, relative to respective parent $PGF_{2\alpha}$ or PGE_2 taken as 1 (mean \pm s.e.).

	Potency			Threshold
		Mean		dose range
Metabolites	Effect	n == 6	Range	μg ml-ĩ
15-ox0	Cons.	1.097	0.36 -1.54	0.008-0.032
PGF1a	Rel.	0.013	0.0025-0.02	0.2 -1.32
13.14-Dihydro	Cons.	1.23	0.47 -1.62	0.02 -0.20
PGF1a		(±0·19)		
13,14-Dihydro	Cons.	0.081	0.0049-0.12	0.02 -0.80
15-0XO PUPA		(±0·0034)		
15-oxo PGE	Rel.	0.16	0.04 -0.25	0.10 -0.40
		(± 0.0074)		
13,14-Dihydro	Rel.	0.12	0.09 -0.16	0.01 -0.20
PGE	D 1	(±0.007)		
13.14-Dihydro	Rel.	0.057	0.04 -0.11	0.10 -0.80
15-oxo PGE		(±0·013)		

ml⁻¹), PPP inhibited the stimulant effect of $PGF_{2\alpha}$, 15-0x0 $PGF_{2\alpha}$, 13,14-dihydro $PGF_{2\alpha}$ and 13,14dihydro 15-0x0 $PGF_{2\alpha}$ on trachea. The contractile effects of histamine and carbacohol were not altered by PPP which itself reduced the resting tone of the tracheal muscle at all concentrations, 0.8 to $400 \,\mu g \, \text{ml}^{-1}$. It was therefore not possible to test its effect on the relaxant effects of PGE_2 and its three metabolites.

Guinea-pig lung resistance in vivo

 $PGF_{2\alpha}$ and 13,14-dihydro $PGF_{2\alpha}$ increased guineapig lung resistance in a dose-dependent manner the latter showing a potency nearly equalling $PGF_{2\alpha}$ (Table 2). The other two metabolites, 15-oxo $PGF_{2\alpha}$ and 13,14-dihydro 15-oxo $PGF_{2\alpha}$ up to 160 μ g kg⁻¹ (higher doses were not used because of the insolubility of the compounds in physiological saline) did not increase lung resistance.

 PGE_2 and its three pulmonary metabolites reduced lung resistance as measured by a reduction in the bronchoconstrictor effects of standard doses of histamine. Responses were dose related. All three metabolites however were weaker bronchodilators when compared with PGE₂ (Table 2).

DISCUSSION

The three metabolites of PGE_2 relaxed guinea-pig tracheal muscle *in vitro* and produced a decrease in lung resistance *in vivo*. However, in both *in vitro* and *in vivo* situations, the metabolites were less active than PGE_2 . The *in vitro* results with the three PGE_2 metabolites are in agreement with those reported by Boot & others (1976) and for 13,14dihydro PGE_2 and 13,14-dihydro 15-oxo PGE_2 the

Table 2. Relative potencies of $PGF_{2\alpha}$ and PGE_2 metabolites on the guinea-pig lung resistance preparation in vivo, relative to respective parent compounds taken as 1 (mean \pm s.e.).

	Potency		otency	Threshold		
		Mean	-	dose range		
Metabolites	Effect	n = 3	Range	µg kg⁻¹		
15-oxo PGF ₁₀		No r	esponse up to 16	0 μg kg‴¹		
13,14-Dihydro	Cons.	0.89	0.59 -1.22	16-62		
PGF ₁ a						
13,14-Dibydro		(±0·13)*				
15-oxo PGF _{1α}	No response up to $160 \ \mu g \ kg^{-1}$ Dil. 0.043 0.017–0.084 0.35–4.0					
15-oxo PGE	Dil.	0.043	0.017-0.084	0-35-4-0		
13.14-Dihydro						
PGE,	Dil.	0.67	0.30 -1.17	0.05-0.5		
13,14-Dihydro						
15-oxo PGE	Dil.	0.028	0.023-0.039	2.0 -4.0		

* n = 4.

results are similar to those reported by Crutchley & Piper (1975). However, for 15-keto PGE₂, Crutchley & Piper (1975) have reported a potency of 0.8-1.8(mean 1.3) compared with PGE₂ whereas in the present study and that of Boot & others (1975) this metabolite is much less active than PGE₂. The effects of the three metabolites of PGE₂ on guineapig lung resistance *in vivo* have not been previously reported. The most potent of the metabolites was 13,14-dihydro PGE₂ which was about as active as PGE₂. Boot, Dawson & Osborne (1976) reported that PGE₂ and 15-oxo PGE₂ relaxed guinea-pig bronchial smooth muscle, the metabolite having similar activity to the parent compound.

Amongst the metabolites of $PGF_{2\alpha}$, 13,14dihydro PGF_{2 α} is a slightly more potent stimulant of guinea-pig tracheal muscle in vitro than $PGF_{2\alpha}$. 13,14-Dihydro 15-oxo $PGF_{2\alpha}$ is approximately 10 times less active than PGF_{2a}. Dawson, Lewis & others (1974) have previously shown that on the guinea-pig trachea, 15-oxo PGF_{2a} was 2-3 times more active than $PGF_{2\alpha}$. I have found 15-oxo $PGF_{2\alpha}$ to be as potent as $PGF_{2\alpha}$ in contracting guinea-pig trachea and also to possess a relaxant component which is seen at a higher concentration. This phenomenon is not unlike that observed for PGE₁, E₂, A₁ and F_{1 α} on isolated small resistance arteries of the rabbit (Strong & Bhor, 1967). In guinea-pig lung experiments in vivo, 13,14-dihydro $PGF_{2\alpha}$ was about as active as $PGF_{2\alpha}$ in increasing lung resistance. The results with 15-oxo $PGF_{2\alpha}$ and 13,14-dihydro $PGF_{2\alpha}$ are in agreement with those reported in the dog by Wasserman (1975).

In the physiological situation, $PGF_{2\alpha}$ is converted to 15-oxo $PGF_{2\alpha}$ by the enzyme 15-hydroxy dehydrogenase and the metabolite is readily reduced to 13,14-dihydro 15-oxo $PGF_{2\alpha}$ by the enzyme

13,14-prostaglandin reductase. Cornette, Harrison & Kirton (1974) reported that the blood concentration of PGF_{2α} in the rhesus monkey is 10-20 times lower than that of 13,14-dihydro 15-000 PGF_{2α} while that of 15-0x0 PGF_{2α} is slightly higher than that of PGF_{2α}. It is therefore likely that in the normal situation, during metabolic degradation, PGF_{2α} is readily transformed into 13,14-dihydro 15-0x0 PGF_{2α} which possesses little biological activity.

There is evidence to suggest that prostaglanding are involved in bronchospasm (Smith, 1976). The relevant effect of 15-oxo PGF_{2α} may relate to the observations of Gardiner & Collier (1974) and Dawson & Boot (1977) who suggest that the response of respiratory muscle to PGs may relate to the initial tone of the tissue although this has not been my experience. Imbalance of PG metabolizing enzymes has been suggested as playing a role in the aetiology of asthma by Dawson & Sweatman (1975), whilst commenting on the biological activity of parent PGs and their metabolites and has also been developed by Mathé (1976), and their present findings add support to these views.

PPP has previously been shown to antagonize some of the effects of PGE_2 and $PGF_{2\alpha}$ (Eakins, 1972). Mathé, Strandberg & Aström (1971) have demonstrated that PPP is capable of blocking the constrictor effect of $PGF_{2\alpha}$ but not the relaxant action of PGE₁ or PGE₂ on isolated human bronchi. On the contrary, Puglisi (1973) reported that PPP antagonized PGE1-induced relaxation but not PGF₂₀-induced contraction on isolated guinea-pig trachea. PPP, however, has not previously been tested for antagonistic action against prostaglandin metabolites on bronchial smooth muscle of any species. In my preparations of guinea-pig trachea in vitro, PPP was shown to block the action of all three PGF₂₀ metabolites. Control doses of carbachol or histamine producing similar responses were not affected by PPP. At the doses tested, PPP consistently produced relaxation of guinea-pig tracheal muscle tone. In this relaxed state it was difficult to observe the antagonistic action of PPP on the relaxant effect of PGE₂ or its three metabolites.

Acknowledgements

All the prostaglandins used in this study were supplied by The Upjohn Company, Kalamazoo, Michigan, U.S.A. The work was carried out under U.S.A.I.D. Grant Contract No. AID/CM/pha-C-73-36 and a research grant from the Lee Foundation, Singapore and the Singapore Turf Club.

REFERENCES

AKCASU, A. (1952). J. Pharm. Pharmac., 4, 671.

- ANGGARD, E. (1965). Biochem. Pharmac., 14, 1507-1516.
- **BOOT**, J. R., DAWSON, W. & HARVEY, J. (1976). Advances in Prostaglandin and Thromboxane Research, Vol. 2. **BOOT**, J. R., DAWSON, W. & HARVEY, J. (1976). Advances in Prostaglandin and Thromboxane Research, Vol. 2. **P** J. R. DAWSON, W. & Construction of the State of
- **BOOT**, J. R., DAWSON, W. & OSBORNE, D. J. (1976). Br. J. Pharmac., 58, 471P.
- CORNETTE, J. C., HARRISON, K. L. & KIRTON, K. T. (1974). Prostaglandins, 5, 155-164.
- CRUTCHLEY, D. J. & PIPER, P. J. (1975). Br. J. Pharmac., 54, 397-399.
- DAWSON, W. & BOOT, J. R. (1977). Fedn Proc. Fedn Am. Socs exp. Biol., 36, 944.
- DAWSON, W., LEWIS, R. L., MCMAHON, R. E. & SURATNAM, W. J. F. (1974). Nature, 250, 331-332.
- DAWSON, W. & SWEATMAN, W. J. (1975). Int. Arch Allergy, 49, 213-216.
- EAKINS, K. E. (1972). Prostaglandins, Progress in Research, pp. 266-292. Editor: Karim, S. M. M. Lancaster: MTP.
- GARDINER, P. J. & COLLIER, H. O. J. (1974). Symposium on Evaluation of Bronchodilator Drugs, Royal College of Physicians, October 1973.
- JAMES, G. W. L. (1969). J. Pharm. Pharmac., 21, 379-386.
- KARIM, S. M. M., SANDLER, M. & WILLIAMS, E. D. (1967). Br. J. Pharmac. Chemother., 31, 640-344.
- MATHÉ, A. A. (1976). Acta physiol. scand., Suppl., 441.
- MATHÉ, A. A., STRANDBERG, K. & ASTRÖM, A. (1971). Nature (New Biol.), 230, 215-216.
- PUGLISI, L. (1973). Adv. Biosci., 9, 219-227.
- SMITH, A. P. (1976). Advances in Prostaglandins Research: Physiological, Pharmacological and Pathological Aspects, pp. 86-103. Editor: Karim, S. M. M., Lancaster: MTP.
- STRONG, C. G. & BOHR, F. (1967). Am. J. Physiol., 213, 725-733.
- SWEATMAN, W. W. & COLLIER, H. O. J. (1968). Nature, 217, 69.
- WASSERMAN, M. A. (1975). Prostaglandins, 9, 959-973.